Neurocranium shape variation of piranhas and pacus (Characiformes: Serrasalmidae) in association with ecology and phylogeny

KELLY S. BOYLE* and ANTHONY HERREL

UMR 7179 C.N.R.S./M.N.H.N., Département d’Ecologie et de Gestion de la Biodiversité, 57 rue Cuvier, Case postale 55, Paris Cedex 5 75231, France

Received 19 March 2018; revised 14 June 2018; accepted for publication 15 June 2018

The teleost neurocranium houses the brain and sense organs, articulates with the upper jaw and suspensorium, and provides the attachment site for muscles involved in feeding, ventilation and locomotion. Serrasalmid fishes (piranhas and pacus) exhibit variation in diet, body shape and habitat that all may influence the evolution of neurocranium shape. In addition, serrasalmids are otophysans with well-developed hearing provided by modified ears and anterior vertebrae that may also influence otic neurocranium morphology. We conducted a 3D geometric morphometrics analysis on high-resolution tomographic data (micro-computed tomography) from 54 museum specimens to measure neurocranium shape variation and disparity among 20 species from the three major serrasalmid clades. We found broad shape variation among species that was explained by phylogenetic history and ecology and related to features such as orbit size and skull depth. Pterotic spine length was longest in carnivorous piranhas and impacts the anatomy of the adductor musculature involved in biting. We found evidence that the otic neurocranium is a separate evolutionary module influenced by both phylogeny and ecology. Lagenar bulla size and position differed among ecological groups and thus sound may vary in importance among species. Small bullae were found in rheophilic herbivores that live among noisy rapids. Thus, serrasalmid neurocranium shape may vary with the relative size of sense organs and with functional aspects of feeding and prey handling.


INTRODUCTION

The neurocranium is an important structure in vertebrates that encloses the brain, houses sensory organs, and provides insertion sites for muscles involved in feeding, ventilation and locomotion (Hanken & Hall, 1993; Hanken & Thorogood, 1993; Helfman et al., 2009). The neurocranium consists of the chondrocranium, formed from cartilaginous capsules that develop around the sense organs of the head, and the dermaticranium, which consists of superficial bones thought to have evolved from scales (Helfman et al., 2009). Teleost fishes exhibit a broad diversity in neurocranium morphology (Gregory, 1933; Alexander, 1964; Arratia, 2003). In addition to phylogenetic patterns, head and neurocranium shape are predicted to be associated with variation in sensory function, feeding adaptations and locomotion (Alexander, 1964; Clabaut et al., 2007; Helfman et al., 2009; Schmitz & Wainwright, 2011b).

Because the neurocranium supports major sensory organs, a strong relationship between morphology associated with sense organs and ecological features of taxa, such as habitat characteristics and prey type, may be predicted. For example, nocturnal fishes are thought to have larger eye diameters to increase sensitivity to low-light conditions for feeding at night (Goatley & Bellwood, 2009; Schmitz & Wainwright, 2011a). Thus, variation in the relative importance of olfaction, vision, hearing, balance and lateral line organs may influence the morphological evolution of the neurocranium. In addition, size-related changes in processing regions of the brain may further influence...
the relative size and shape of the cranium (Striedter & Northcutt, 2006).

For aquatic animals, the shape of the cranium may be especially important for locomotion because of the density of the surrounding medium and the drag forces experienced during locomotion (Segall et al., 2016). There may be a relative trade-off between drag and the relative depth of the head and the ability to manoeuvre and make rapid turns (Webb, 1984, 2005). Furthermore, the shape of the head is related to overall body depth (Alexander, 1964), a feature that may reduce the number of potential predators by gape limitation (Nilsson & Brönmark, 2000; Claverie & Wainwright, 2014).

Neurocranium shape can be expected to be influenced by functional demands influencing the evolution of muscles involved in locomotion, feeding and ventilation. For example, the supraoccipital forms the anterior origin of the epaxial musculature of the body (Alexander, 1964; Winterbottom, 1974). Moreover, some adductor mandibulae fibres, involved in closing the lower jaw and biting, originate on portions of the neurocranium (Alexander, 1964; Winterbottom, 1974; Datovo & Castro, 2012). Thus, neurocranium shape variation may influence the size and position of the origins of cranial muscles, which would be expected to have functional consequences. In addition, neurocranium shape variation influences the position of other cranial bones, which could influence the position and size of cranial muscles and the biomechanics of feeding and ventilation. For example, neurocranium shape influences the position of the suspensorium that articulates with the lower jaw, position of the upper jaw, opercular series and gill arches.

The shape of the otic region of the neurocranium can be expected to be related to the function of the ears in hearing and balance. Changes in size and shape of otic bones may be associated with size differences of the semicircular canals and otolithic end organs of the ear. The relative sizes of otoliths and sensory maculae vary substantially in fishes (Popper & Fay, 2011). Among otophysan fishes, the relative sizes of the asteriscus otolith and associated lagena end organ are substantially larger than in most teleosts, while the sagitta and sacculus are relatively reduced in size (Popper & Fay, 2011). The lagener ooliths lie within large lateral capsules of the neurocranium in otophysans (Frost, 1925). The lagena of otophysan fishes is thought to be involved in the detection of sound particle motion, while the sacculus functions principally in the role of indirect detection of sound pressure stimuli that are transduced by the Weberian apparatus (Fay, 1984; Popper & Fay, 2011). The Weberian apparatus morphology (Weber, 1820) is unique to otophysan fishes and involves a chain of ossicles connected by ligaments and associated with the anterior vertebrae. This morphology is associated with additional modifications of the otic region of the skull, in which a medial opening, the cavum sinus impar, exists ventral to the foramen magnum. Sound pressure-induced vibrations of the swim bladder are mechanically transduced to the anterior Weberian ossicles (scaphiae). The cavum sinus impar allows fluid motion between the scaphiae and a perilymphatic space (sinus impar) in the inner ears of otophysan fishes (Alexander, 1962; Popper & Fay, 1973; Chardon & Vandewalle, 1997). As a result of this morphology, otophysan fishes are able to hear higher frequencies of sound pressure in comparison to most other teleosts (von Frisch, 1938; Popper, 1972; Ladich, 1999; Yan et al., 2000; Mélotte et al., 2018).

Serrasalmid fishes (piranhas and pacus) provide an opportunity to examine variation of the neurocranium among species that vary in feeding ecology, feeding behaviour and phylogenetic history. This characiform family is composed of approximately 93 species (Eschmeyer & Fong, 2017) and 16 extant genera (Thompson et al., 2014) with a broad distribution in South America from Atlantic coast river drainages to the eastern slopes of the Andes. The family includes fruit-eating and seed-dispersing species (Goulding, 1980; Goulding & Carvalho, 1982; Horn et al., 2011) and carnivorous species with diets that vary in relative importance and include whole prey of invertebrates and fishes and pieces of larger fishes (fins and scales) (Machado-Allison & Garcia, 1986; Nico & Taphorn, 1988; Sazima & Machado, 1990; Nico, 1991). Several rheophilic herbivore species of serrasalmids are restricted to high-energy river environments with torrents and rapids and feed extensively on submerged aquatic plants of the family Podostemaceae (Jégu & Dos Santos, 2002; Jégu et al., 2002a, b; Jégu & Keith, 2005; Andrade et al., 2013). In addition, serrasalmid body shapes range from moderately slender to relatively deep-bodied. Thus, the neurocranial morphology of serrasalmid species may be influenced by a variety of factors: phylogenetic history, feeding morphology, body shape and sensory morphology. Variation in habitat and feeding behaviour may require a larger role of certain sensory modalities, such as vision and hearing. For example, there is anecdotal evidence that splash sounds associated with falling seeds and fruits may be attractive to some fruit-eating serrasalmids (Gottsberger, 1978; Piedade et al., 2006; Correa et al., 2007; Parolin et al., 2010) and carnivorous piranhas are reported to be attracted to splashing sounds from wounded prey and the feeding activities of other piranhas (Markl, 1972; Stabentheiner, 1988; Mol, 2006). In contrast, rheophilic herbivorous serrasalmids probably live in a noisier environment and their diet of submerged aquatic plants may not be associated with an acoustic stimulus.
Serrasalmid neurocranium morphology is predicted to be influenced by both phylogenetic history and ecology. In addition, feeding ecology in serrasalmids is influenced by phylogenetic history (K. S. Boyle & A. Herrel, unpubl. obs.). We tested five hypotheses. (1) Neurocranium morphology is associated with phylogeny and (2) neurocranium morphology is partly explained by ecology, even after accounting for phylogenetic history. In addition, because hearing may be especially relevant to the ecology of otophysan fishes, and because we predict different sound cues to be relevant to specific serrasalmid feeding ecologies, we expected a relationship between ecology and otic neurocranium morphology. We tested hypothesis (3) that the otic neurocranium has evolved as a modular structure relative to the rest of the neurocranium, (4) that otic neurocranium morphology is associated with phylogeny and (5) that otic neurocranium morphology is partly explained by ecology after

Figure 1. Landmarks used in analyses to quantify shape variation of the left side of the neurocranium. Reconstructions come from *Serrasalmus elongatus* Kner, 1858 MNHN IC.1989.1360 (left in each pane) and *Myleus setiger* (Müller & Troschel, 1844) MNHN IC.1998.1156 (right). A, lateral view; B, ventral view; C, dorsal view; D, oblique dorsal anterior view. Numbered red dots correspond to landmarks (see Table 1 for definitions). Black dots represent sliding semi-landmarks on curves. Blue dots represent sliding surface landmarks.
accounting for phylogenetic history. We tested for allometry as an alternative explanation of shape variation of the neurocranium and otic neurocranium. We also examined areas of pattern-based morphological convergence of species within and among ecological groups that may explain neurocranium morphological variation among serrasalmids. In addition, we tested for differences in disparity among ecological groups and among the three major clades: piranha, Myleus-pacus and pacu (sensu Ortí et al., 2008; Thompson et al., 2014).

MATERIAL AND METHODS

PHyLOGENY

A composite phylogeny (Supporting Information, Fig. S1) was constructed based on data from molecular phylogenies (Ortí et al., 2008; Thompson et al., 2014; Andrade et al., 2017) that together included 18 of the 20 species in our study. Our study included two species that were not present in available phylogenetic hypotheses: Metynnis lippincottianus (Cope, 1870) and Serrasalmus elongatus Kner, 1858. We inferred the phylogenetic position of these two taxa based on taxonomy and placed M. lippincottianus as sister to its only congener in our dataset, M. hypsauchen (Müller & Troschel, 1844), and we placed S. elongatus as sister to S. rhombeus (Linnaeus, 1766) in a nexus file made based on the Thompson et al. (2014) phylogeny. To construct the composite phylogeny, we used the matrix with rate parsimony (mrp) method (Sanderson et al., 1998). Phylogeny nexus files were transformed into a ‘multiphylo’ object with the ‘c.phylo’ function of the ‘APE’ library (Paradis et al., 2004) in R (R Core Team, 2017). The composite tree was produced using the ‘pratchet’ method and 10 000 iterations using the ‘mrp.supertree’ function of ‘phytools’ R library (Revell, 2017). We used branch length estimates from Thompson et al. (2014) when available. When branch length estimates were not available, we assigned branch length values by evenly dividing evolutionary time of the most recent calibrated node among the remaining branches (Aquino & Colli, 2017).

SPECIMENS, μCT SCANS AND 3D RECONSTRUCTIONS

Specimens from the study were obtained from the Ichthyology Collection at the Muséum National d’Histoire Naturelle (MNHN), Paris, France (Table S1). We used micro-computed tomography (μCT) scanning to produce 3D surfaces of the neurocranium (Figs 1, 2). Specimens were scanned at the AST-RX technical platform at the MNHN with a μCT scanner (v|tome|x 240 L, GE Sensing & Inspection Technologies phoenix|x-ray). Voxel sizes ranged from 31.1 to 100.0 μm (mean 53.6). Segmentation and

Figure 2. Landmarks used in analyses to quantify shape variation of the left side of the neurocranium. Reconstructions come from Serrasalmus elongatus MNHN IC.1989.1360 (left in each pane) and Myleus setiger MNHN IC.1998.1156 (right). A, anterior view; B, posterior view; C, oblique dorsal posterior view. Numbered red dots correspond to landmarks (see Table 1 for definitions). Black dots represent sliding semi-landmarks on curves. Blue dots represent sliding surface landmarks.
surface generation were performed using AVIZO 7.0 (FEI, Mérignac, France). Semi-automatic thresholding was used to identify bone. A drawing tablet was used to manually isolate the bones of the neurocranium from the other bones of the skull, the supraneural of the neural complex, first vertebra, scaphium, posttemporal bones and the otoliths. The remaining segmented bones were used to create a singular bilateral neurocranium surface file that consisted of the vomer, mesethmoid, lateral ethmoids, frontals, parasphenoid, orbitosphenoid, parietales, occipitales, supraoccipitales, sphenotics, pterotics, prootics, basioccipital, exoccipitales and intercalars. Surface generation was performed using the 'constrained smoothing' setting in AVIZO 7.0. Surfaces were exported as (.PLY) format files. Surface files were decimated to reduce file size (between 440 176 and 1 607 463 triangles) and cleaned to fill in holes, remove creased edges remove and spikes with

<table>
<thead>
<tr>
<th>Landmark</th>
<th>Definition</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Anterior tip of mesethmoid</td>
</tr>
<tr>
<td>2</td>
<td>Anteriolateral corner of vomer</td>
</tr>
<tr>
<td>3</td>
<td>Posteriolateral corner of vomer</td>
</tr>
<tr>
<td>4</td>
<td>Ventral tip of the lateral ethmoid</td>
</tr>
<tr>
<td>5</td>
<td>Suture of lateral ethmoid and frontal along lateral edge</td>
</tr>
<tr>
<td>6</td>
<td>Anterior or superior corner of the orbitosphenoid (depends on orientation of bone in different specimens)</td>
</tr>
<tr>
<td>7</td>
<td>Posterior or inferior corner of the orbitosphenoid (depends on orientation of bone in different specimens)</td>
</tr>
<tr>
<td>8</td>
<td>Suture of frontal and sphenotic, along lateral edge</td>
</tr>
<tr>
<td>9</td>
<td>Ventral tip of sphenotic</td>
</tr>
<tr>
<td>10</td>
<td>Inferior corner of pterotic spine</td>
</tr>
<tr>
<td>11</td>
<td>Tip of pterotic spine</td>
</tr>
<tr>
<td>12</td>
<td>Superior corner of pterotic spine</td>
</tr>
<tr>
<td>13</td>
<td>Intersection of the parietal, supraoccipital and epioccipital at the posterior margin of the parietal</td>
</tr>
<tr>
<td>14</td>
<td>Tip of the epioccipital</td>
</tr>
<tr>
<td>15</td>
<td>Suture between parietal and supraoccipital along the dorsal ridge</td>
</tr>
<tr>
<td>16</td>
<td>Tip of supraoccipital crest</td>
</tr>
<tr>
<td>17</td>
<td>Centre of mesethmoid, posterioremend</td>
</tr>
<tr>
<td>18</td>
<td>Anterior medial corner of frontal bone along the frontal fonatenelle</td>
</tr>
<tr>
<td>19</td>
<td>Left, anterior intersection of epiphyseal bar and frontal</td>
</tr>
<tr>
<td>20</td>
<td>Left, posterior intersection of epiphyseal bar and frontal</td>
</tr>
<tr>
<td>21</td>
<td>Suture between the parietal and frontal along the frontal fontanelle</td>
</tr>
<tr>
<td>22</td>
<td>Anterior ventral corner of exoccipital, just superior to the basioccipital</td>
</tr>
<tr>
<td>23</td>
<td>Anterior dorsal corner of basioccipital, just inferior to the exoccipital</td>
</tr>
<tr>
<td>24</td>
<td>Posterior edge of the lagenar bulla and exoccipital</td>
</tr>
<tr>
<td>25</td>
<td>Posterior edge of the lagenar bulla and basioccipital</td>
</tr>
<tr>
<td>26</td>
<td>Junction of anterior end of parasphenoid and vomer</td>
</tr>
<tr>
<td>27</td>
<td>Ventral tip of the parasphenoid, below orbit, crux below the posterior margin of the orbit, anterior to the lamellae that fuse to the basioccipital</td>
</tr>
<tr>
<td>28</td>
<td>Posterior end of left basioccipital lamellae</td>
</tr>
<tr>
<td>29</td>
<td>Ventral medial point of vertebral articulation on basioccipital</td>
</tr>
<tr>
<td>30</td>
<td>Posteroventral tip of supraoccipital</td>
</tr>
<tr>
<td>31</td>
<td>Anterior portion of exoccipital that meets a strut along the edge of the foramen magnum</td>
</tr>
<tr>
<td>32</td>
<td>Dorsal medial point of vertebral articulation on basioccipital</td>
</tr>
<tr>
<td>33</td>
<td>Centre of vertebral articulation on basioccipital</td>
</tr>
<tr>
<td>34</td>
<td>The medial edge of the anterior end of the left exoccipital bone that lies over the sinus endolymphaticus</td>
</tr>
<tr>
<td>35</td>
<td>The medial edge of the posterior portion of the exoccipital, superior to the sinus impar</td>
</tr>
<tr>
<td>36</td>
<td>The medial point of the posterior opening of the cavity sinus impar, part of exoccipital</td>
</tr>
<tr>
<td>37</td>
<td>Lateral corner of nVII foramen in prootic</td>
</tr>
<tr>
<td>38</td>
<td>Medial corner of nVII foramen in prootic</td>
</tr>
<tr>
<td>39</td>
<td>Anterior corner of the prootic ‘auditory’ foramen</td>
</tr>
<tr>
<td>40</td>
<td>Posterior corner of the prootic ‘auditory’ foramen</td>
</tr>
</tbody>
</table>

*Landmarks used in the subset of data of the auditory neurocranium.
Geomagic Studio software (Geomagic Studio; Raindrop Geomagic, Research Triangle Park, NC, USA). We analysed shapes of the left side of the neurocranium (left elements of paired bones and the left portion from the midline of unpaired elements). Several specimens (Table S1) had irregularities on the left side in which the auditory foramen, nVII foramen or both were not fully ossified and were open to the margin of the prootic and one specimen had a damaged left lagenar bulla. These specimens were mirrored in Geomagic Studio and the right side was used for analysis.

ECOLOGICAL GROUPS

Fish species were categorized into three broad ecological groups for analysis based on published accounts of feeding habits and habitat associations (Table S1). These ecological groups were: (1) carnivores, species that mainly consume fish prey either whole or in pieces (muscle tissue, lepidophagy, fins); (2) frugivores/omnivores, species that consume a substantial portion of fruits and seeds; and (3) rheophilic herbivores, species that are restricted in habitat to high-flow environments among rapids and waterfalls and that have a largely herbivorous diet composed of submerged plants. We distinguished rheophilic herbivores and omnivorous frugivores/omnivores species because these species are more restricted in habitat to high-energy environments and feed mainly on submerged associated plants associated with these habitats, such as species in the family Podostemaceae (Jégu et al., 2002a, b; Jégu & Keith, 2006; Dary et al., 2017). Other serrasalmid species in our study can occur among rapids (e.g. Fitzgerald et al., 2018), but have broader habitat associations and differ in feeding ecology (Table S1). Acnodon normani Gosline, 1951, a Brazilian shield species with a broad diet that includes fruits, seeds, leaves, insects and scales (Leite & Jégu, 1990), was included in the frugivore/omnivore group. This species tends to occur in slow to moderate flowing waters, relative to more rheophilic serrasalmids (Zuanon, 1999). Ancestral state reconstruction indicates that a frugivore/omnivore ecology is the ancestral condition for the species in our dataset (K. S. Boyle & A. Herrel, unpubl. obs.) and occurs in 11 of 20 species in our study, including all members of the pacu clade, most members of the Myleus-pacu clade, and within the genus Metynnis of the piranha clade. Ancestral state reconstruction indicates that rheophilic herbivory, which in our dataset occurs as a monophyletic lineage with the Myleus-pacu clade, probably evolved once (K. S. Boyle & A. Herrel, unpubl. obs.). Carnivores occur as a derived monophyletic clade within the piranha clade and probably evolved once (K. S. Boyle & A. Herrel, unpubl. obs.).

GEOMETRIC MORPHOMETRIC LANDMARKS

Forty homologous landmarks (Table 1) were identified and placed on the left (l.) side of the neurocranium (Fig. 2). Homologous landmarks were placed on the mesethmoid, vomer, l. lateral ethmoid, l. frontal, l. orbitosphenoid, l. sphenotic, l. pterotic, l. parietal, supraoccipital, l. epicapital, epiophyseal bar, l. exoccipital, basioccipital and parasphenoid of surface files in IDAV Landmark software (Wiley et al., 2005). We used a combination of traditional homologous landmarks and sliding 3D semi-landmarks with the goal of providing a better description of structures devoid of good homologous landmarks (Bookstein, 1997; Gunz et al., 2005). Sliding semi-landmarks are geometrically homologous landmarks that can be used to increase the resolution to describe shapes (383 landmarks in total). Curve semi-landmarks were allowed to slide along a trajectory between defined homologous landmarks and surface semi-landmarks were allowed to slide across surfaces. Curve semi-landmarks were placed along curve features between homologous landmarks using the Landmark software. A total of 21 curves with 8–20 semi-landmarks per curve was used (280 semi-landmarks in total). Curves were placed along the left side of the frontal fontanelle from the supraoccipital crest; to the anterior end of the fontanelle; around the left margin of the mesethmoid; along the ventral midline of the paraphenoid; along the left margin of the orbit; around the left pterotic spine; along the caudal midline of the supraoccipital; along the curve between the supraoccipital, parietal and epicapital on the left side; and around the left side of the basioccipital at its articulation with vertebra 1 (Fig. 1). In addition, several curves associated with the otic region of the neurocranium were chosen (Fig. 1): a curve along the ventral margin of the exoccipital along the dorsal portion of the left lagenar bulla; a curve along the dorsal margin of the basioccipital along the ventral portion of the left lagenar bulla; a curve around the left side to the midline of the posterior opening of the cavum sinus impar; a curve along the left-medial edge of the dorsal surface of exoccipital bone within the brain case that covers the sinus endolymphaticus of the ears; a curve around a foramen in the prootic for a ramus of nVII that is open to the utricle; and a curve around the prootic foramen, a characiform synapomorphy (Fink & Fink, 1981) that is also open to the utricle. To generate semi-landmarks, a single specimen for each bone was arbitrarily chosen as an initial template. Surface landmarks were added to the template using Landmark software and used to project surface landmarks on to all the specimens for each bone or otolith using the ’placePatch’ function of the ’Morpho’ library (Schlager, 2015) in R (R Core Team, 2017). Eleven surface landmarks were placed on the l. exoccipital over the dorsal,
outer portion of the lagenar bulla, 18 on the basioccipital on the lower outer portion of the left lagenar bulla, seven on the left side of the supraoccipital crest, three on the dorsal surface of the l. sphenotic, six on the dorsal surface of the l. frontal, two on the parasphenoid, three on the vomer, three on the l. lateral ethmoid, two on the mesethmoid and four on the left side of the ephiphysyeal bar. Curve and semi-landmarks were slid using ‘Morpho’ (Schlager, 2015) to minimize the bending energy of the thin plate spline, first between each specimen and the template (over five iterations), and then between each specimen and the Procrustes consensus of the entire dataset over ten iterations (Gunz et al., 2005; Botton-Divet et al., 2015).

Subsequent analyses were done on the full left neurocranium 383 landmark dataset as well as a subset of landmarks that characterizes the otic region of the skull. This subset included ten of the original homologous landmarks (Table 1) from the sliding procedure described above, and the curves on the exoccipital and basioccipital over the lagenar bulla, the curves around the nVII foramen and prootic foramen, the curve over the dorsal roof of the sinus endolymphaticus, the curve around the left side of the exoccipital’s articulation with vertebra 1, and the curve around the left side of the cavum sinus impar. This subset of landmarks included the surface landmarks of the dorsal and ventral portions of the left lagenar bulla. This subset of otic neurocranium data included a total of 320 landmarks (landmarks, curve semi-landmarks and surface landmarks).

Generalized Procrustes superimpositions (Rohlf & Slice, 1990) of the entire data set and for the auditory neurocranium subset were conducted in the ‘geomorph’ library (Adams et al., 2016) in R (R Core Team, 2017) using the ‘gpagen’ function. For this Procrustes superimposition, the previously slid curve and surface semi-landmarks were treated as fixed landmarks. For each dataset, the mean shape of each species was calculated using the ‘mshape’ function of ‘geomorph’ in order to allow for phylogenetic comparisons among species. A second Procrustes superimposition on the species means was then calculated for each structure as described above using the ‘gpagen’ function of ‘geomorph’ for subsequent analyses. Principal components analysis (PCA) was conducted on the species means using ‘PlotTangentSpace’ in ‘geomorph’. Phylogenetic relationships with the PCA morphospace were visualized with the ‘phylomorphospace’ function in the ‘phytools’ library (Revell, 2017) in R (R Core Team, 2017). To produce a species-mean neurocranium and otic neurocranium surface file to the mean shape. We used the ‘tps3d’ function in ‘Morpho’ (Schlager, 2015) to produce superimposed representations of the shape of neurocrania and otic neurocrania on the extremes of PC axes.

**Phylogenetic signal**

We estimated phylogenetic signal in shape data with the multivariate K-statistic (Adams, 2014) using the Procrustes coordinate data for the full neurocranium and for auditory neurocranium datasets. These tests were applied using the ‘physignal’ function of ‘geomorph’ (Adams et al., 2016). In addition, we examined phylogenetic signal present on the individual PC axes that contributed to at least 75% of the cumulative variation with a univariate lambda test (Ives et al., 2007) with the ‘phylosig’ function in the ‘phytools’ library (Revell, 2012).

**Modularity of the otic neurocranium**

We tested for modularity of the otic region of the neurocranium relative to the other neurocranium landmarks. We used the covariance ratio (CR) test of Adams (2016). This test measures the degree of independence between structures (modularity) by examining the covariance within and among modules. We conducted this test with the ‘modularity.test’ function in ‘geomorph’ (Adams et al., 2016) and used 999 iterations to assess significance by randomization of landmark subsets between modules. In addition, to account for potential effects of phylogenetic history on the degree of observed modularity among species, we performed the same test under a Brownian motion model of evolution using the ‘phylo.modularity’ function in ‘geomorph’ (Adams et al., 2016).

**Shape allometry**

We performed several tests to determine if the shape relationships among species in our study were influenced by size. Using the ‘stats’ package in R, we performed Pearson correlation tests between the log10-transformed means of centroid size for the neurocranium and otic neurocranium and the PC axes that explained at least 75% of the shape variation. In addition, for both the neurocranium and the otic neurocranium, we performed a Procrustes regression test for patterns of shape covariation among species using the ‘procD.allometry’ function of ‘geomorph’ (Adams et al., 2016). These models used the natural log transformation of mean centroid size for each species and initially included an interaction with ecological
group, which was dropped from the model if not significant. Lastly, we considered phylogeny in tests of size covariation with shape. We used the `procD.pglsl' function of 'geomorph' (Adams et al., 2016) to perform phylogenetic generalized least squares (PGLS) Procrustes regressions for the neurocranium and otic neurocranium. We tested models with and without an ecological group interaction term and used the natural log of mean centroid size for each species.

SHAPE CONVERGENCE

To test for putative cases of convergent evolution of neurocranium shape, we used the approach of Stayton (2017) to test for pattern-based similarity by quantifying the amount of phenotypic distance between lineages closed by evolution. After examination of phylomorphospace plots from PCA and between-groups PCA (bgPCA), we identified two examples of putative convergence. We used the 'convratsig' function of the 'convervol' package (Stayton 2017) in R to calculate the proportion of the maximum phenotypic distance between putatively convergent taxa closed by evolution (C), the magnitude of phenotypic distance closed by evolution (C), the proportion of the magnitude of convergence relative to the phenotypic distance of the lineages being compared (C), and the proportion of the magnitude of convergence relative to the phenotypic distance of all lineages descended from the most recent common ancestor of the putatively convergent taxa (C) (Stayton, 2017). For both cases of putatively convergent taxa, we tested convergence among total neurocranium shape and total otic neurocranium shape by using PCA scores (all axes that together described at least 75% of variation) to calculate phenotypic distance with 'convervol'. In addition, we performed two additional tests of convergence of the shape features associated with the greatest separation among ecological groups for the entire neurocranium and otic neurocranium using the scores from the bgPCAs. To calculate statistical significance of measures of convergence (C), we performed 1000 simulations.

SHAPE–ECOLOGY RELATIONSHIPS

We tested for relationships between overall shape of the neurocranium and the auditory neurocranium and ecology using phylogenetic MANOVA tests (Garland et al., 1993) on the PC axes that collectively explained ≥ 75% of variation. These tests were conducted with the 'aov.phyl' function of the 'geiger' library (Harmon et al., 2015) in R (R Core Team, 2017). For these tests, we ran 1000 simulations and used Wilk’s lambda as our multivariate test. In cases where differences were detected among groups (P < 0.05) from the 1000 simulations, we used the 'phylANOVA' function to perform phylogenetic ANOVA followed by Holm’s post-hoc test using the 'phytools' package.

We used bgPCA to determine which subset of shape features best discriminate the a priori-defined ecological groups. This method produces orthogonal axes and avoids the problems of small within-group sample sizes and high-dimensional geometric morphometric data for analyses such as Canonical Variates Analysis (Mitteroecker & Bookstein, 2011). The bgPCA uses the co-variance matrix of the means of a priori-defined groups, and then projects the individual data on to the eigenspace defined by the between-group covariance matrix (Mitteroecker & Bookstein, 2011; Schlager, 2015). Two bgPCAs were performed, one on the entire neurocranium landmark dataset, and one on the subset of landmarks associated with the auditory region of the skull. To perform these analyses, we used the ‘groupPCA’ function of the ‘Morpho’ library (Schlager, 2015). Species scores from the two axes of the bgPCA were plotted as a phylomorphospace using the ‘phylomorphospace’ function in the ‘phytools’ library (Revell, 2017) in R (R Core Team, 2017). We used the ‘tps3d’ function in ‘Morpho’ (Schlager, 2015) to produce superimposed representations of the shape of neurocrania and otic neurocrania on the extremes of bgPC axes. Differences among the Euclidean distances of the bgPCA scores from the three groups were tested by permutation (10,000 rounds) in ‘Morpho’. We also tested for differences among bgPCA scores of the three groups while accounting for phylogeny with a phylogenetic MANOVA and for post-hoc comparisons used phylogenetic ANOVA and Holm’s tests as described above.

MORPHOLOGICAL DISPARITY AMONG ECOLOGICAL GROUPS

Morphological disparity was measured for the entire neurocranium landmark dataset and the subset of landmarks from the auditory region. To perform these analyses, we used the ‘morphol.disparity’ function of the ‘geomorph’ library (Adams et al., 2016). The Procrustes variance estimate is a measure of the sum of the diagonal elements of the group covariance matrix divided by sample size (Adams et al., 2016).

RESULTS

OVERALL NEUROCRANION SHAPE

The first three principal components (PCs) explained 52, 14 and 10% of the variation in neurocranium shape and the first five PCs explained 90% of the cumulative variation (Fig. 3, Table 2).
Phylogenetic signal was evident on the first three PCs (Fig. 3) as revealed by the univariate $\lambda$ (Table 2). In addition, the multivariate $K$-statistic indicated significant phylogenetic signal for overall neurocranium shape ($K_{\text{alt}} = 0.82, P = 0.001$).

No evidence of allometry of neurocranium shape among species was observed. Correlations of centroid size of neurocranium landmarks and PC scores among species were weak (Table 2). Procrustes regression indicated no shape covariation with size among species ($P_{1,18} = 0.582, P = 0.702, R^2 = 0.031$) and no interaction between size and ecological group was observed ($P = 0.05$). Similarly, after accounting for phylogeny with PGLS regression, no relationship between size and shape was observed ($P_{1,18} = 1.035, P = 0.230, R^2 = 0.068$) and no interaction with size and ecology was observed ($P > 0.05$).

PC1 varied largely with respect to the relative height of the supraoccipital crest, length of the snout/mesethmoid region, size and anterior position of the orbit, length of the pterotic spine, and angle of the parasphenoid (Fig. 3). Both ecological groups and members of the three major serrasalmid clades were widely distributed along PC1 (Fig. 3A, B). Carnivores and frugivore/granivores displayed wide variation in shape features associated with PC1. For example, the relative elongate skull of the carnivore *Serrasalmus elongatus* Kner, 1858, which has a short supraoccipital crest, had the lowest score on PC1, while the carnivore *Captrion mento* (Cuvier, 1819), which is relatively deep and has a tall supraoccipital crest, had the highest score among carnivores and the third highest score of all taxa on PC1 (Fig. 3A). PC2 varied largely with respect to the relative size of the orbit, anterior and lateral extent of the orbit, and relative size of the lagenar bulla (Fig. 3). High scores on PC2, associated with anteriorly positioned eyes and large lagenar bullae, separated carnivores from rphiletic herbivores, while frugivore/granivores were broadly distributed along PC2 (Fig. 3A, E). Major serrasalmid clades were largely distinguished along PC2, with low scores associated with *Mylossoma*–pacus, moderate scores associated with piranha clade members, and high scores associated with the pacu clade (Fig. 3B). High scores on PC3 were associated with a large pterotic spine and low scores were associated with a curved parasphenoid that descends ventrally (Fig. 3E). Carnivores were distinguished from rphiletic herbivores and most frugivore/granivores by high scores on PC3 (Fig. 3C). Major serrasalmid clades were distributed broadly across PC3 (Fig. 3D). For example, *Metynnis hypsauchen* (Müller & Troschel, 1844) and *M. lippincottianus* (Cope, 1870) had lower scores than the other members of the piranha clade, which were probably associated with their short pterotic spines.

PCA revealed (Fig. 3) overall shape similarity between *C. mento* of the piranha clade and *Mylossoma asterias* (Müller & Troschel, 1844), *Mylossoma rubripinnis* (Müller & Troschel, 1844) and *Mylossoma schomburgkii* (Jardine, 1844) of the *Mylossoma*–pacus clade. This observation was supported by measures of convergence higher than expected by chance ($C_1 = 0.481, P < 0.001; C_2 = 0.079, P < 0.001; C_3 = 0.049, P < 0.001; C_4 = 0.059, P < 0.001$). Both lineages are characterized by a deep skull, with high supraoccipital crest (Fig. S3). *Captrion mento*, however, is characterized by a longer, straighter pterotic spine (Fig. S2). *Metynnis lippincottianus* and *M. hypsauchen* showed shape similarity with *Mylossoma aureum* (Spix & Agassiz, 1829) and *M. duriventris* (Cuvier, 1818) on PC1 and PC3 (Fig. 3). These two genera had a higher $C_1$ measure of convergence than expected by chance ($C_1 = 0.291, P = 0.019$), but other measures of convergence were not statistically significant ($C_2 = 0.19, P = 0.262; C_2 = 0.307; C_4 = 0.012, P = 0.307$). Both genera are characterized by a slightly deeper skull and tall supraoccipital crest than average and a shorter pterotic spine (Fig. S3). *Metynnis*, however, differs from *Mylossoma* in having more dorsally orientated parasphenoid (Fig. S4). By contrast, *C. mento* diverged in shape from the elongate skull of *Pristobrycon striolatus* and *Serrasalmus elongatus*, and *Acnodon normani* was divergent in shape from other *Mylossoma*–pacus in having an elongate skull and large orbits (Fig. 3A, B).

**ECOLOGICAL GROUP RELATIONSHIPS TO OVERALL NEUROCRANIUM SHAPE**

Phylogenetic MANOVA performed on PCs 1–3 indicated differences among ecological groups (Wilks $\lambda = 0.12, F_{2,17} = 9.40, P_{\text{non-phylo}} < 0.001, P_{\text{phylo}} = 0.024$). Phylogenetic ANOVA indicated a difference on PC3 between carnivore and frugivore/granivores ($F_{2,17} = 13.75, P_{\text{non-phylo}} < 0.001, P_{\text{phylo}} = 0.036$, Holmes post-hoc test $P = 0.048$), but not between rphiletic herbivores and carnivores ($P = 0.214$).

Between-group PCA of the overall neurocranium shape indicated that carnivores were best distinguished from frugivore/granivores along bgPC1 and were characterized by having a more anteriorly positioned orbit, a straight parasphenoid with an orientation mainly along the rostro-caudal axis, an elongate pterotic spine and lagenar bullae in a more dorsal position (Fig. 4). High scores on bgPC1 were also associated with a smaller supraoccipital crest (Fig. S3). Carnivores were best distinguished from rphiletic herbivores by having a more dorsally orientated parasphenoid (Fig. S2). Carnivores were characterized by having a more anteriorly positioned eye and large lagenar bullae, separated carnivores from rphiletic herbivores, while frugivore/granivores were broadly distributed along PC2 (Fig. 3A, E). Major serrasalmid clades were largely distinguished along PC2, with low scores associated with *Mylossoma*–pacus, moderate scores associated with piranha clade members, and high scores associated with the pacu clade (Fig. 3B). High scores on PC3 were associated with a large pterotic spine and low scores were associated with a curved parasphenoid that descends ventrally (Fig. 3E). Carnivores were distinguished from rphiletic herbivores and most frugivore/granivores by high scores on PC3 (Fig. 3C). Major serrasalmid clades were distributed broadly across PC3 (Fig. 3D). For example, *Metynnis hypsauchen* (Müller & Troschel, 1844) and *M. lippincottianus* (Cope, 1870) had lower scores than the other members of the piranha clade, which were probably associated with their short pterotic spines.
Figure 3. Overall shape variation of serrasalmid neurocrania revealed by principal components analysis (PCA). Phylomorphospace biplots of (A, B) principal components (PCs) 1 and 2 and (C, D) PCs 1 and 3. A, phylomorphospace with ecological groups outlined with minimum convex polygons (green = frugivore/granivores, blue = rheophilic herbivores, red = carnivores) shows broad overlap of frugivore/granivores and other groups on PCs 1 and 2, while rheophilic herbivores are distinguished from carnivores by lower scores on PC2. B, biplot of PCs 1 and 2 with three major serrasalmid clades outlined (yellow = pacu clade, light blue = Myluses-pacu clade, pink = piranha clade) shows overlap of all three clades on PC1, but more separation at the clade level on PC2. C, carnivores are separated from rheophilic herbivores and most frugivore/granivores along PC3. D, wide variation exists among the piranha clade along PC3, while most members of the pacus clade are distinguished from Myluses-pacus and piranha clade members by low scores on PC3. E, superimposed neurocranial surfaces indicate the shapes associated with the extremes of the PC axes; purple shapes = positive extremes, orange shapes = negative extremes. The mean shape of all species in the study is shown on the left in panel E. Top left, lateral view (anterior to the right); top right, caudal view (lateral to the left); middle, dorsal view (anterior to the right); bottom, ventral view (anterior to the right).
along bgPC2, while frugivore/granivores displayed wide variation along this axis (Fig. 4). High scores on bgPC2 were characterized by a more anteriorly directed nasal region, larger and more posteriorly positioned orbits, a supraoccipital crest with less dorsal extension and extended more broadly posteriorly, and small lagenar bullae (Fig. 4). *Acnodon normani*, which has large orbits and a neurocranium that is less deep than most serrasalmids, had the highest scores on bgPC2, despite having large lagenar bullae. Permutation tests on the Euclidean distances of bgPCA scores indicated significant differences between carnivores and frugivore/granivores (*P* = 0.018) and between carnivores and rheophilic herbivores (*P* = 0.038), but not between rheophilic herbivores and frugivore/granivores (*P* = 0.41). However, differences among bgPCA scores of ecological groups were not evident after accounting for phylogeny (phyMANOVA Wilks $\lambda = 0.27$, $F_{2,17} = 7.35$, *P* non-phylo < 0.001, *P* phylo = 0.089).

As was observed for the overall neurocranium morphology identified by the standard PCA, *C. mento* showed convergence in neurocranium morphology identified by bgPCA to *M. asterias*, *M. rubripinnis* and *M. schomburgkii*. Measures of convergence were higher than expected by chance ($C_1 = 0.511$, $P = 0.003$; $C_2 = 0.063$, $P = 0.001$; $C_4 = 0.060$, *P* < 0.001; $C_5 = 0.070$, *P* < 0.001). In addition, *Metynnis* and *Mylossoma* had a higher $C_1$ measure of convergence than expected by chance ($C_1 = 0.464$, $P = 0.004$), but other measures of convergence were not statistically significant ($C_2 = 0.029$, $P = 0.091$; $C_3 = 0.028$, $P = 0.071$; $C_4 = 0.028$, $P = 0.071$).

### Overall Neurocranium Shape Disparity

Carnivores had the greatest overall neurocranium shape disparity (Procrustes variance = 27.49 × 10$^{-3}$), followed by frugivore/granivores (19.20 × 10$^{-3}$) and rheophilic herbivores (8.33 × 10$^{-3}$). The overall observed variation (sum of group covariance matrix diagonal elements) was highest for carnivores (0.1100), followed by frugivore/granivores (0.0768) and rheophilic herbivores (0.0333). A permutation test did not reveal significant differences in disparity among ecological groups (*P* > 0.05 for all comparisons). Among the three clades, the piranha clade had the greatest shape disparity (22.73 × 10$^{-3}$, observed variation 0.0909), followed by the *Myleus*-pacu clade (17.99 × 10$^{-3}$, observed variation 0.0719) and the pacu clade (15.24 × 10$^{-3}$, observed variation 0.0609). A permutation test did not reveal statistically significant differences in disparity among clades (*P* > 0.05 for all comparisons).

### Shape of the Otic Neurocranium

The otic and non-otic regions of the neurocranium showed a significant degree of modularity. CR was 0.955 (*P* = 0.001) for the non-phylogenetic model and 0.977 (*P* = 0.001) for the phylogenetic model.

The first three PCs from a PCA performed on a subset of neurocranium landmarks of the otic region explained 46, 16 and 9% of the variation in neurocranium shape and the first five PCs explained 85% of the cumulative variation (Fig. 5, Table 2).

<table>
<thead>
<tr>
<th>Structure</th>
<th>PC</th>
<th>Variation (%)</th>
<th>Cumulative %</th>
<th>Univariate phylogenetic signal</th>
<th>Pearson’s product-moment correlation between centroid size of species and PC score</th>
</tr>
</thead>
<tbody>
<tr>
<td>Neurocranium</td>
<td>PC1</td>
<td>52.1</td>
<td>52.1</td>
<td>1.011 0.037</td>
<td>0.23 0.98 18 0.340</td>
</tr>
<tr>
<td></td>
<td>PC2</td>
<td>14.3</td>
<td>66.4</td>
<td>1.108 &lt; 0.001</td>
<td>0.25 1.12 18 0.278</td>
</tr>
<tr>
<td></td>
<td>PC3</td>
<td>10.1</td>
<td>76.5</td>
<td>1.026 &lt; 0.001</td>
<td>−0.10 −0.44 18 0.669</td>
</tr>
<tr>
<td></td>
<td>PC4</td>
<td>9.4</td>
<td>85.9</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td>PC5</td>
<td>3.9</td>
<td>89.9</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Neurocranium</td>
<td>PC1</td>
<td>45.9</td>
<td>45.9</td>
<td>1.101 0.005</td>
<td>0.31 1.38 18 0.186</td>
</tr>
<tr>
<td>Otic</td>
<td>PC2</td>
<td>16.0</td>
<td>62.0</td>
<td>1.015 &lt; 0.001</td>
<td>−0.19 −0.83 18 0.417</td>
</tr>
<tr>
<td>Region</td>
<td>PC3</td>
<td>9.2</td>
<td>71.2</td>
<td>1.017 0.021</td>
<td>0.25 1.07 18 0.296</td>
</tr>
<tr>
<td></td>
<td>PC4</td>
<td>7.3</td>
<td>78.5</td>
<td>0.261 0.540</td>
<td>−0.38 −1.77 18 0.094</td>
</tr>
<tr>
<td></td>
<td>PC5</td>
<td>6.1</td>
<td>84.6</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Table 2. Summary of the variation of principal component (PC) analyses on the geometric morphometric data, univariate lambda tests of phylogenetic signal conducted on individual PC axes, and correlations between mean centroid size and shape (PC score) on each axis.

Bold type indicates statistical significance (*P* < 0.05).
indicated significant phylogenetic signal ($K_{\text{mult}} = 0.75$, $P = 0.001$).

No evidence of allometry of otic neurocranium shape among species was observed. Correlations of centroid size of otic neurocranium landmarks and PC scores among species were weak (Table 2). Procrustes regression indicated no shape covariation with size among species ($F_{1,18} = 1.00$, $P = 0.372$, $R^2 = 0.053$) and

**Figure 4.** A, between-group principal components analysis (bgPCA) to determine the features of the overall neurocranium landmark dataset that best distinguish ecological groups. Carnivores (red polygons) are distinguished from rheophilic herbivores (blue) by low scores on bgPC2, while frugivore/granivores (green) overlapped in morphospace with carnivores and rheophilic herbivores on bgPCs 1 and 2. B, superimposed neurocrania surfaces indicate the shapes associated with the extremes of the bgPC axes: purple shapes = positive extremes, orange shapes = negative extremes. The mean shape of all species in the study is shown on the left in panel B. Left, lateral view (anterior to the right); middle, caudal view (lateral to the left); top right, dorsal view (anterior to the right); bottom right, ventral view (anterior to the right).
no interaction between size and ecological group was observed ($P = 0.05$). After accounting for phylogeny with PGLS regression, no relationship between size and shape among species was observed ($F_{1,18} = 1.22$, $P = 0.280$, $R^2 = 0.064$). However, an interaction between ecological group and size was observed ($F_{1,2,14} = 2.25$, $P = 0.007$, $R^2 = 0.205$). This interaction appears to be influenced by shape divergence between carnivores and rheophilic herbivores that have otic neurocranial of similar size and broad shape and size variation of the otic neurocrania of frugivore/granivores (Fig. S4).

High scores on PC1 were associated with a relatively small lagenar bulla, a large vertebral articulation of the basioccipital, and anteriorly positioned prootic and nVII foramina (Fig. 5A, E). Frugivore/granivores were widely distributed along PC1, while carnivores and rheophilic herbivores were less distributed along this axis of shape space and did not overlap with each other (Fig. 5A). Of the major serrasalmid clades, the Myleyus-pacus clade was broadly distributed along PC1, while the piranha and pacu clades occupied more narrow, and largely non-overlapping regions along PC1 (Fig. 5B).

Low scores on PC2 were associated with more posteriorly positioned lagenar bullae and anteriorly positioned prootic and nVII foramina, and a more anteriorly positioned opening of the cavum sinus impar (Fig. 5E). All ecological groups overlapped on PC2, with rheophilic herbivores tending to have higher PC2 scores than carnivores (Fig. 5A). Major serrasalmid clades overlapped along the PC2 shape axis, with the highest scores exhibited within the Myleyus-pacus clade (Fig. 5B). Low scores on PC3 were associated with larger lagenar bullae, a smaller and rounder vertebral articulation surface of the basioccipital, and posteriorly positioned prootic and nVII foramina (Fig. 5E).

Rheophilic herbivores had higher scores than carnivores along PC3, while frugivores/granivores were broadly distributed along this axis (Fig. 5C). All three major serrasalmid clades were broadly distributed and largely overlapping along PC3 (Fig. 5D).

In contrast to the pattern observed for the entire neurocranium, C. mento did not display shape convergence of the otic neurocranium with $M. asterias$, $M. rubripinnis$ and $M. schomburgkii$ ($C_1 = 0.069$, $P = 0.440$; $C_2 = 0.006$, $P = 0.488$; $C_3 = 0.004$, $P = 0.539$; $C_4 = 0.006$, $P = 0.519$). *Catoprion mento* is characterized by an expanded lagenar bulla and a longer prootic with a greater distance between the lagenar bulla and the prootic and nVII foramina relative to these Myleyus-pacus species (Fig. S2). Convergence of otic neurocranium morphology between Metynnis and Mylossoma was observed, as was seen previously for the overall neurocranium ($C_1 = 0.301$, $P = 0.005$; $C_2 = 0.037$, $P = 0.007$; $C_3 = 0.026$, $P = 0.010$; $C_4 = 0.026$, $P = 0.010$). Both Metynnis and Mylossoma are characterized by a larger lagenar bulla and more anteriorly positioned prootic foramen and nVII foramen (Fig. S3).

**ECOLOGICAL GROUP RELATIONSHIPS TO OTIC NEUROCRANIUM SHAPE**

A MANOVA performed on the first four PCs indicated differences among ecological groups (Wilks $\lambda = 0.21$, $F_{2,17} = 4.07$, $P_{\text{non-phylo}} = 0.003$), but these differences were not significant after accounting for phylogenetic history (phyMANOVA, $P_{\text{phylo}} = 0.361$). Differences were also not observed on individual PC axes (1–4), however, when tested with ANOVA ($P > 0.05$).

Between-group PCA of the otic region of the neurocranium indicated that carnivores were best distinguished from rheophilic herbivores along bgPC1 and were characterized by larger lagenar bullae that extended more ventrally, a smaller and rounder vertebral articulation of the basioccipital, a taller opening of the cavum sinus impar, and a prootic foramen that is positioned more caudally (Fig. 6). Frugivore/granivores were best distinguished from carnivores and rheophilic herbivores by lower scores along bgPC2 and were characterized by having the largest lagenar bullae, a shallower opening of the cavum sinus impar and a prootic foramen that was wider laterally (Fig. 6). Permutation tests on the Euclidean distances of bgPCA scores indicated significant differences between carnivores and rheophilic herbivores ($P = 0.022$), but not between carnivores and frugivore/granivores ($P = 0.272$) or between rheophilic herbivores and frugivore/granivores ($P = 0.056$).

Differences in the auditory region of the neurocranium were observed after accounting for phylogeny (Wilks $\lambda = 0.09$, $F_{2,17} = 18.57$, $P_{\text{non-phylo}} < 0.001$, $P_{\text{phylo}} = 0.002$). Differences along bgPC1 were not observed after accounting for phylogeny (phyANOVA $F_{1,17} = 7.84$, $P_{\text{non-phylo}} = 0.004$, $P_{\text{phylo}} = 0.139$). Shape differences were, however, evident along bgPC2 after accounting for phylogeny (phyANOVA $F_{1,17} = 29.56$, $P_{\text{non-phylo}} < 0.001$, $P_{\text{phylo}} = 0.004$): frugivore/granivores differed from rheophilic herbivores and carnivores (Holme’s post-hoc test $P = 0.018$ for both).

*Catoprion mento* did not display shape convergence of otic neurocranium morphology identified by bgPCA with $M. asterias$, $M. rubripinnis$ and $M. schomburgkii$ ($C_1 = 0.204$, $P = 0.248$; $C_2 = 0.011$, $P = 0.376$; $C_3 = 0.013$, $P = 0.408$; $C_4 = 0.017$, $P = 0.388$). *Metynnis* and *Mylossoma*, however, were convergent in otic neurocranium morphology identified from bgPCA ($C_1 = 0.375$, $P = 0.023$; $C_2 = 0.030$, $P = 0.032$; $C_3 = 0.036$, $P = 0.030$; $C_4 = 0.036$, $P = 0.030$).
Figure 5. Shape variation of the otic region of serrasalmid neurocrania revealed by PCA. Phylomorphospace biplots of (A, B) principal components (PCs) 1 and 2 and (C, D) PCs 1 and 3. A, phylomorphospace with ecological groups outlined with minimum convex polygons (green = frugivore/granivores, blue = rheophilic herbivores, red = carnivores) shows broad overlap of frugivore/granivores with rheophilic herbivores and carnivores on PCs 1 and 2, while carnivores are distinguished from rheophilic herbivores on PC1, but not PC2. B, biplot of PCs 1 and 2 with three major serrasalmid clades outlined (yellow = pacu clade, light blue = Myleus-pacu clade, pink = piranha clade) shows that members of the pacu clade are largely distinguished from piranha clade members along PC1, while pacu clade members are largely distinguished from Myleus-pacu clade members along PC2. C, frugivore/granivores are widely distributed along PC3 and overlap both carnivores and rheophilic herbivores. Carnivores are distinguished from rheophilic herbivores by lower scores on PC3. D, all three major serrasalmid clades show broad overlap along PC3. E, superimposed otic neurocranium region surfaces indicate the shapes associated with the extremes of the PC axes: purple shapes = positive extremes, orange shapes = negative extremes. The mean shape of all species in the study is shown on the left in panel E. The rendered surface includes portions of the prootic, exoccipital and basioccipital, while the parietal, frontal and supraoccipital have been removed. Top left, dorsal view of the
Otic neurocranium shape disparity

Frugivore/granivores had the greatest shape disparity (Procrustes variance = 14.28 × 10⁻³, observed variation 0.0571), followed by rheophilic herbivores (9.95 × 10⁻³, observed variation 0.0398) and carnivores (9.83 × 10⁻³, observed variation 0.0393). A permutation test did not reveal statistically significant differences in disparity among clades (P > 0.05 for all comparisons). Among the three clades, the pacu clade had the greatest shape disparity (15.20 × 10⁻³, observed variation 0.0608) followed by the Myleus-pacus (13.03 × 10⁻³, observed variation 0.0521) and the piranha clade (9.72 × 10⁻³, observed variation 0.0389). A permutation test did not reveal statistically significant differences in disparity among clades (P > 0.05 for all comparisons).

DISCUSSION

This study demonstrated that serrasalmid neurocranium morphology is influenced by both phylogeny and ecology and that some shape aspects are associated with ecology after accounting for evolutionary history. In addition, the otic and non-otic neurocranium are modular and the otic neurocranium module is influenced by phylogeny and ecology after accounting for phylogeny. Neurocranium shape variation is associated principally with the depth of the skull, the size and position of the orbit, the length of the pterotic spine, and the relative size of the lagenar bulla. This observed variation in neurocranium shape was not explained by variation in size and no evidence of shape allometry was observed. Dorsal elongation of the skull, resulting from expansion of the supraoccipital crest and ventral expansion of the parapophyses, has evolved repeatedly within the Serrasalmidae, occurring in each of the three major clades and the frugivore/granivore and carnivore ecological groups. Several areas of observed variation are probably correlated with sensory roles and may be related to ecology: elongation of the nasal region, position and size of the orbit, and relative size of the lagenar bulla. Evidence of modularity between otic and non-otic neurocranium regions indicates that the function of the ear may have influenced the shape evolution of this region independently relative to other regions of the neurocranium. The restricted analysis of the otic region of the neurocranium indicated strong differences among ecological groups associated with the size of the lagenar bulla, the relative position of the bulla with regard to the basisphenoid – vertebra 1 articulation, and the rostro-caudal position of the two foramina of the prootic. Lastly, there is a strong phylogenetic trend associated with some aspects of skull shape: laterally expanded orbits in the piranha and pacu clade, an elongate pterotic spine as a derived feature within the piranha clade, and anteriorly positioned lagenar bullae in Myleus-pacus clade members.

In this study, a major axis of shape variation related to depth and elongation of the skull was detected. This main axis of variation has been observed in other studies of head shape in fishes, for example in populations of three-spine stickleback Gasterosteus aculeatus (Pistore et al., 2016). Relatively deep skulls were associated with a taller supraoccipital crest. Elaboration of the supraoccipital crest appears to be related to body depth in serrasalmids (Alexander, 1964). The increased height of vertebrae, neural spines and associated epaxial musculature requires a firm, large origin, as provided by the supraoccipital crest (Alexander, 1964). In this study, convergent deep neurocranium morphologies were observed between the lepidophagous carnivore Catoprion mento and several frugivore/granivores in the Myleus-pacus clade. However, these species differed in the length and shape of the pterotic spine and morphology of the otic neurocranium. Convergence in deep-bodied neurocrania was also observed in Mylossoma. In this study, deep skulls were often associated with a parasphe- noid with a greater dorso-ventral extent. Serrasalmids show variation in body depth within each of the three major clades. Relatively elongate-bodied species occurred in the piranha clade (Serrasalmus elongatus, Pristobrycon striolatus and S. rhombeus) and in the Myleus-pacus clade (Acnodon normani). The degree of body depth vs. elongation is a major source of repeated evolutionary variation among teleost fishes (Ward & Brainerd, 2007; Friedman, 2010; Sallan & Friedman, 2012; Claverie & Wainwright, 2014). Among other distantly related teleost lineages that vary in relative body length vs. depth, e.g. Anabantoides, elongation of the head and skull is a main source of overall elongation (Collar et al., 2016). This study found multiple examples of divergence between sister taxa along the axis of neurocranium depth and elongation: C. mento vs. P. striolatus, Acnodon normani vs. derived Myleus-pacus, and Colossoma macropomum.

Elongation of the neurocranium of serrasalmids in this study involved either extension of the nasal/ethmoid region or elongation of the otic region of the skull, the latter of which resulted in a relatively more anterior position of the eyes on the head. Anteriorly positioned orbits were characteristic of piranhas in this
Figure 6. Between-group principal components analysis (bgPCA) to determine the features of the otic region of the neurocranium landmark dataset that best distinguish ecological groups. Carnivores (red polygons) are distinguished from rheophilic herbivores (blue) by low scores on bgPC2, while frugivore/granivores (green) overlapped in morphospace with carnivores and rheophilic herbivores on bgPCs 1 and 2. Superimposed neurocrania surfaces indicate the shapes associated with the extremes of the PC axes; purple shapes = positive extremes, orange shapes = negative extremes. Left, lateral view (anterior to the right); middle, caudal view (lateral to the left); top right, dorsal view (anterior to the right); bottom right, ventral view (anterior to the right). Superimposed otic neurocranium region surfaces indicate the shapes associated with the extremes of the PC axes; purple shapes = positive extremes, orange shapes = negative extremes. The mean shape of all species in the study is shown on the left in panel E. The rendered surface includes portions of the prootic, exoccipital and basioccipital, while the parietal, frontal and supraoccipital have been removed. Top left, dorsal view of the bone overlying the sinus endolymphaticus (anterior is to the right); middle, caudal oblique dorsal view that shows the left portion of the cavum sinus impar (lateral to the left); right, caudal view (lateral to the left); bottom left, ventro-lateral view of the lagenar bulla and prootic with prootic and nVII foramina visible (anterior is to the right).
NEUROCRANIUM SHAPE VARIATION OF PIRANHAS AND PACUS

study and may be predicted to be associated with the role of vision in the predatory habits of these species. Orbit position also affects the location of the suspensorium and hyomandibula articulation behind the orbit, and the location of the pterotic. These two features have ramifications for feeding because they affect the position of the lower jaw articulation and the length and angle of adductor mandibulae muscle fibres.

Several serrasalmids in this study were characterized by relatively large orbits. Eye size is correlated with visual acuity and habitat type (Caves et al., 2017). *Acodon normani* has a large orbit that is positioned at a large relative distance to the anterior end of the neurocranium. In addition, the cranium is relatively narrow in lateral aspect in this species. Piranha orbits also tended to be large, but, as mentioned above, were located further anteriorly.

One source of variation among serrasalmid skulls was the relative length of the pterotic spine. As noted by Alexander (1964), the pterotic spine reaches the level of the supracleithrum in piranhas. A long pterotic spine is a feature that was observed in all carnivores of the piranha clade and members of the pacu clade, especially *Colossoma macropomum* (Cuvier, 1816) and *Piaractus brachypomus* (Cuvier, 1818). A relatively short pterotic spine was present among most of the other frugivore/granivores in this study, including the frugivore/granivore members of the piranha clade (*Metynnis* spp.), as well as the rheophilic herbivores. The pterotic spine is the origin for some muscle fibres involved in closing the jaws. Some fibres of the pars malaris (originally A2) subdivision of the adductor mandibulae originate on the pterotic and pterotic spine of *Serrasalmus* (Alexander, 1964; Mirande, 2009; Datovo & Castro, 2012; Datovo & Vari, 2014). In *Serrasalmus* the pterotic spine provides part of the origin of the A2 subdivision of the adductor mandibulae (Alexander, 1964). Some A2 lateral fibres also originate on the pterotic spine (Datovo & Castro, 2012). Thus, future studies should examine if variation in the elaboration of this bone is associated with feeding and biting.

The orientation of the parasphenoid varies among serrasalmid species, from a near-horizontal orientation along the rostro-caudal axis, exhibited in species such as *Serrasalmus elongatus*, to an orientation that extends ventrally from rostral to caudal in *Metynnis* spp. and *Myleus* spp. Parasphenoid orientation also varied between taxa with neurocrania that were convergent in other aspects (*Metynnis* and *Myleosoma*). Variation in this orientation is apparent in the first two principal components of overall neurocranium shape (Fig. 3) and in the first axis of the analysis of overall shape between ecological groups (Fig. 4). Orientation of this bone may have implications in biting and prey processing. In *Serrasalmus* spp., some fibres of the A2 subdivision of the adductor mandibulae originate on the parasphenoid and pterosphenoid, and some fibres of the A3 adductor mandibulae originate on the parasphenoid, along with the pterosphenoid and metapterygoid (Datovo & Castro, 2012). In addition, in serrasalmids the parasphenoid is the origin of the adductor arcus palatini, which inserts on the dorso-medial face of the hyomandibula (Datovo & Castro, 2012). This muscle may be involved in compression of the buccal cavity during feeding and swallowing, as was observed in distantly related serrasalmids (Lauder, 1981). Future studies should examine if orientation of the parasphenoid in serrasalmids accompanies variation in muscle fibre length and orientation in the adductor mandibulae and adductor arcus palatini that may influence bite force and prey transport, respectively. In addition, orientation and length may have ramifications for neurocranium strength during feeding.

**OTIC NEUROCRANIUM**

In this study, the otic neurocranium varied according to both ecology and phylogeny. The observed modularity between otic and non-otic neurocranium regions is consistent with the hypothesis that hearing and vestibular sensory evolution has influenced the morphological evolution of the neurocranium. Patterns of convergence of the overall neurocranium and otic neurocranium are in agreement with the observed modularity. For example, convergence between *C. mento* and several *Myleus* pacu clade members was apparent for the overall neurocranium in general, but not for the otic neurocranium. Serrasalmids, like most otophysans, are characterized by acute hearing and sound pressure sensitivity that exists because of the presence of a Weberian apparatus (Stabenheiner, 1988; Ladich, 1999; Mélotte et al., 2018). In addition, serrasalmids have well-developed lagener otoliths (asterisci), which in otophysan fishes are relatively large and associated with particle motion detection (Fay, 1984; Popper & Fay, 2011). Variation in the size of the lagener bulla among ecological groups indicates that the importance of sound particle motion detection may vary with ecology. Small lagener bullae in this study were associated with rheophilic herbivorous ecologies. These differences parallel differences in relative asteriscus size and shape differences observed in rheophilic herbivore serrasalmids (K. S. Boyle & A. Herrel, unpubl. obs.; K. S. Boyle et al., unpubl. obs.). Sound particle motion has a greater relative impact in the acoustic nearfield, which extends far in aquatic environments at lower frequencies (Bass & Clark, 2003). Splash sounds that incorporate low-frequency elements may be important
particle motion stimuli for both frugivores/granivores and carnivores. Splash sounds from falling seeds and fruits are thought to be attractive to some serrasalmids (Gottsb erg, 1978; Piedad et al., 2006; Correa et al., 2007; Parolin et al., 2010). Splashing, which can occur from wounded prey and from the swimming activities of piranhas feeding, is attractive to piranhas (Markl, 1972; Stabentheiner, 1988; Mol, 2006). The small lagenar bullae of rheophilic herbivores indicates that particle motion detection may be less important. The feeding ecology of these species is herbivory of submerged aquatic plants (Podostemaceae) (Jégud & Dos Santos, 2002; Jégud et al., 2002b; Jégud & Keith, 2005; Orti et al., 2008; Andrade et al., 2013; Dary et al., 2017). Sounds are not expected to be associated with this food resource and it is not known if much sound is produced by rheophilic herbivores feeding on these plants. Furthermore, rheophilic habitats are predicted to be noisy environments (Lugli & Fine, 2003, 2007; Tonolla et al., 2011) and perhaps high levels of particle motion background noise would select for a smaller lagena and lagenar bulla. Smaller otoliths are predicted to respond differently than larger otoliths to sound, with larger otoliths likely to have a greater response to low frequencies (Lychakov & Rebane, 2000; Popper et al., 2005). An alternative hypothesis is that rheophilic environments favour particle motion sensitivity at higher frequencies, and thus a smaller asteriscus, which does not require a large lagenar bulla. In addition, relative size differences of the asteriscus and surrounding lagenar bulla of rheophilic herbivores compared to other serrasalmids may be related to differences in balance and postural requirements of life in high-energy environments such as rapids. The otolithic end organs of teleost fishes are hypothesized to be multifunctional by providing both vestibular and hearing senses (Popper & Platt, 1993). A negative relationship is predicted to exist between otolith size and balance requirements in fishes, and is supported, in part, by the small otolith sizes of large pelagic fishes that are capable of rapid angular accelerations (Popper et al., 2005). Serrasalmids in high-energy rheophilic environments may need to manoeuvre more rapidly in currents in eddies. Consistent with this hypothesis, the lapillus of the utricule of rheophilic herbivore serrasalmids has a smaller relative size than in frugivore/granivore species (K. S. Boyle & A. Herrel, unpubl. obs.). Importantly, hypotheses regarding the function of lagenar bulla reduction related to particle motion hearing and balance requirements of rheophilic herbivorous lifestyles are not mutually exclusive.

A prootic foramen that is open to the utriculus is a synapomorphy of characiform fishes (Fink & Fink, 1981). In addition, a foramen providing passage for a ramus of the facial nerve exists on the prootic, anterior to the prootic foramen, which also exposes the utriculus. It is not known if these structures contribute to hearing or the degree to which the utriculus is involved in hearing in serrasalmids. Variation in the size and position of these foramina along the rostrocaudal axis was observed in this study. For example, the prootic foramen tended to be wider among frugivore/granivores than in the other ecological groups. These foramina were in a relatively anterior position in Colossoma macropomum and Piaractus brachypomus and were in a relatively posterior position in Acnodon normani. Notably, these foramina occur in a more intermediate position in Mylossoma aureum and M. duriventre relative to the closely related C. macropomum and P. brachypomus. Anterior swim bladder diverticulae are present in Mylossoma spp. (Nelson, 1961). Moreover, some of these diverticulae approach moderately close to the prootic foramen (K. S. Boyle & A. Herrel, unpubl. obs.). Swim bladder extensions in other fishes can provide increased sensitivity to sound pressure (Yan et al., 2000; Parmentier et al., 2011; Tricas & Boyle, 2015). However, this function seems less plausible for an otophysan fish that already has sensitivity to sound pressure via the Weberian apparatus. Hearing thresholds have not yet been tested for serrasalmids of the genus Mylossoma and are needed to better understand this question.

The fluid-filled sinus connection between the Weberian ossicle linkage and ear, the cavum sinus impar, showed some variation among serrasalmids in this study. Frugivore/granivores tended to have a cavum sinus impar that was in a lower, more shallow position above the occiput. A functional hypothesis for this observed difference is not clear, but shape and position variation of this feature may be expected to co-vary with shape and size differences of the scaphia, which are in close association with this structure.

**Neurocranium Shape Disparity**

Differences in neurocranium disparity among ecological groups were modest, with carnivores displaying the greatest amount of shape disparity in this study, while rheophilic herbivores had the least disparity in overall neurocranium morphology. Disparity among the major clades was also comparable. The oldest lineage, the pacu clade, showed the least disparity. Note, however, that branch lengths to the most recent common ancestor among the three major clades are similar (Fig. S1). Pacu clade members have deep skulls, with long pterotic spines, and large lagenar bullae. Greater shape disparity in the piranha clade could be explained, in part, by the variation in body depth among carnivorous species, and the variation in pterotic spine length between carnivores and the frugivore/granivore species of Metynnis. Intermediate disparity in the Mylossus-pacu clade can be accounted for by variation in skull elongation (e.g. elongate *Acnodon normani*, deep *Myloplus schomburgkii*).
and lagenar bulla size between rheophilic herbivores and frugivore/granivores in this clade. Contrasting patterns of disparity for other cranial and anterior skeletal shape features have been observed in other characiform fishes (Sidlauskas, 2008). In clades of comparable diversity, the more trophically diverse clade shows higher disparity as a result of the morphospace occupied by the tips of the phylogeny (Sidlauskas, 2008). In this study, the clade with the least disparity (pacu clade) was also the only clade with only one ecological group among its members, although this pattern was not observed for otic neurocranium diversity.

Otic neurocranium disparity among ecological groups and clades was also comparable. Carnivores showed the least otic neurocranium disparity (comparable to rheophilic herbivores), while frugivore/granivores showed the greatest disparity. Carnivores had consistently large lagenar bullae and little variation in the position of the prootic and nVII foramina. Frugivore/granivores displayed more variation in the placement of the prootic and nVII foramina. This variation was evident within the pacu clade, which showed more variation in the position of these foramina. The Mylæus-pacu clade showed moderate variation in otic neurocranium disparity, in part because of the variation in lagenar bulla size between rheophilic herbivores and frugivore/granivore members of this clade.

CONCLUSION

This study examined three-dimensional neurocranium shape in serrasalmids. Shape variation of the skull exhibits strong phylogenetic signal and also variation in regions that may be predicted to have consequences for sensory biology, hydrodynamic and body shape functions, and jaw suspension and feeding. The otic region of the neurocranium exhibited differences among ecological groups that parallels observations of otolith size and shape differences in recent studies in serrasalmids. Morphological disparity was not dramatically different among ecological groups or the three major clades of the family, however. Furthermore, the pattern of disparity among ecological groups and clades was not consistent between the overall neurocranium landmark data and the subset of the otic region. These results suggest that patterns of variation of the overall neurocranium may not be parallel to patterns of variation of individual modules, such as the otic neurocranium. This is evident among the carnivorous piranhas, which show broad variation in skull elongation, but a conserved pattern of large lagenar bullae in the otic neurocranium. Future research is needed to determine the consequences of the observed skull shape variation on the functional morphology of locomotion, feeding and sensory ecology in these fishes.

ACKNOWLEDGEMENTS

We thank Zora Gabsi, Philippe Keith and Patrice Pruvost (MNHN, CNRS UMR 7208) for assistance and access to MNHN specimen loans. We are grateful to Miguel Garcia Sanz and Patricia Wils from the AST-RX platform at MNHN (CNRS UMS 2700) for μCT scanning and training in Avizo. We thank four anonymous reviewers for their suggestions that improved the manuscript. This study was funded by a Marie-Skłodowska Curie fellowship (EU project 625039 – EvoMorphASIS).

REFERENCES


**SUPPORTING INFORMATION**

Additional Supporting Information may be found in the online version of this article at the publisher’s web-site.

**Table S1.** Details of the specimens used in study.

**Figure S1.** Phylogenetic hypothesis of serrasalmid species used in our study. Composite tree constructed from Ortí et al. (2008), Thompson et al. (2014) and Andrade et al. (2017). Branch length data are in millions of years (ma). Ecological groups of each species are indicated on the right. Nodes 1, 2 and 3 indicate the ‘pacu clade’, ‘*Myleus*-pacus clade’ and ‘piranha clade’, respectively (sensu Ortí et al., 2008; Thompson et al. 2014).

**Figure S2.** Superimposed landmarks (LMs) of *Catoprion mento* (red LMs), the mean shape of several *Myleus*-pacus (*Myleus asterias, Myloplus rubripinnis, Mylosesinus schomburgkii*; purple LMs), and the mean shape of all 20 serrasalmid species in the study (orange LMs). Mean LM positions were calculated using the ‘mshape’ function of the R package ‘geomorph’ (Adams et al., 2016). *Catoprion mento* LMs are the mean of individuals and the other means (several *Myleus*-pacus, and the dataset grand mean) are means of species means. Superimposed LMs were generated with the ‘deformGrid3d’ function in the R package ‘Morpho’ (Schlager, 2015). *Catoprion mento* vs. the mean shape of several *Myleus*-pacus (A, D, G, J, M), *Catoprion mento* vs. the mean shape of all serrasalmids in the study (B, E, H, K, N), and the mean shape of several *Myleus*-pacus vs. the mean shape of all serrasalmids in the study (C, F, I, L, O). Lateral view (A–C), dorsal view (D–F), ventral view (G–I), caudal view (J–L) and frontal view (M–O).

**Figure S3.** Superimposed landmarks (LMs) of *Mylossoma* (black LMs), *Metynnis* (grey LMs), and the mean shape of all 20 serrasalmid species in the study (orange LMs). Mean LM positions were calculated using the ‘mshape’ function of the R package ‘geomorph’ (Adams et al., 2016). *Mylossoma* LMs are the mean of species means (*M. aureum* and *M. duriventre*), *Metynnis* LMs are the mean of species means (*M. hypsauchen* and *M. lippocotianus*), and the dataset grand mean is the mean of all 20 species means in the study. Superimposed LMs were generated with the ‘deformGrid3d’ function in the R package ‘Morpho’ (Schlager, 2015). *Mylossoma* vs. *Metynnis* (A, D, G, J, M), *Mylossoma* vs. the mean shape of all serrasalmids in the study (B, E, H, K, N), and *Metynnis* vs. the mean shape of all serrasalmids in the study (C, F, I, L, O). Lateral view (A–C), dorsal view (D–F), ventral view (G–I), caudal view (J–L) and frontal view (M–O).

**Figure S4.** Relationship between centroid size (ln centroid size) and shape (PC1) for the otic neurocranium. Carnivores are shown in red, rheophilic herbivores in blue and frugivore/granivores in green.